

Translating the complex immune system into disease insights with CyTOF technology



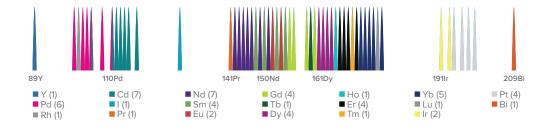
#### Reasons to choose CyTOF

# THE CYTOF

#### Reasons to take your high-dimensional

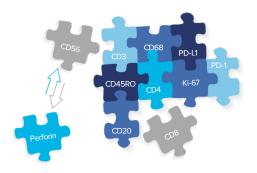
#### Precision data with discrete signals

Not impacted by spectral overlapping of fluorochromes and tissue autofluorescence



#### Easy panel design to complete experiments quicker

Large number of available antibodies without overlap simplifies panel design and expansion



Start with ready-to-go panels and easily swap markers in and out.

## B

#### **Reproducible and comparable**

Stained samples can be frozen, stored and shipped to support longitudinal studies and multi-site workflows.

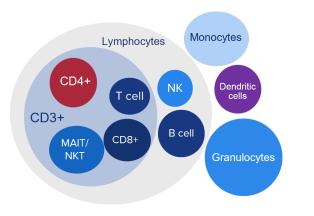


## ADVANTAGE

#### research beyond the limits of fluorescence

#### Capture rare or unexpected cell populations

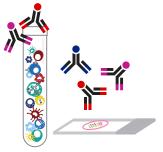
Unbiased, high-dimensional profiling of 40-plus markers to uncover diverse immune subpopulations





#### Minimal sample required, saving on limited clinical research material

Simultaneous staining and detection from a single tube or tissue scan, without multiple staining controls or time-consuming cyclic protocols



## 6

#### **Trusted by researchers**

The leading technology for high-parameter immune research





#### Cytometry by time-of-flight (CyTOF<sup>®</sup> technology)

Applies purified heavy metal labels, not normally found in biological systems, instead of fluorophores



#### Risks of fluorescence

for high-parameter studies



**Missed** cell populations or **false positives** 



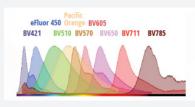




**Reduced sensitivity** where fluorescence overlap occurs



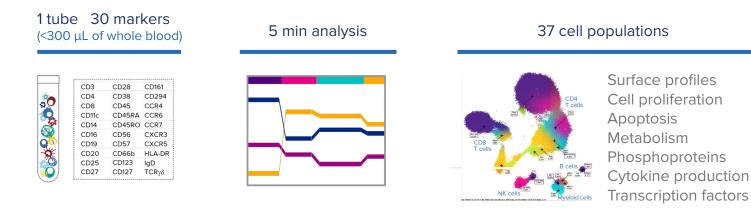
**Higher resource use** to compensate for spectral overlap



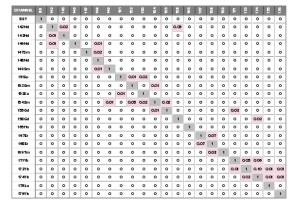
# From sample collection to high-dimensional insights **in 3 days**\*

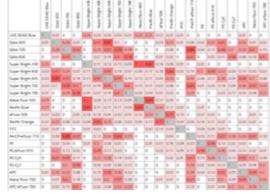
#### **Flow cytometry**

Get started with the validated Maxpar<sup>®</sup> Direct<sup>™</sup> Immune Profiling Assay<sup>™</sup>.



#### Minimal signal overlap





The CyTOF flow cytometry image (far left) shows minimal spillover between metal channels when compared with the same panel from a competitor spectral flow cytometer (left).

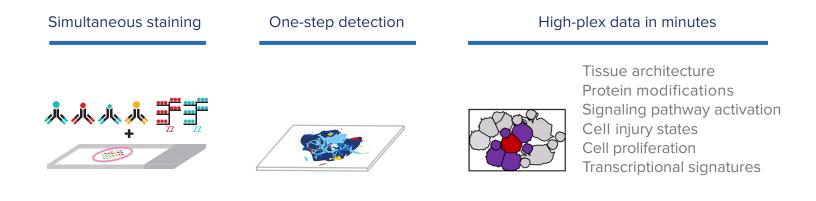
#### Quantify and visualize 40-plus markers in a single run.

\* After panel and image analysis optimization

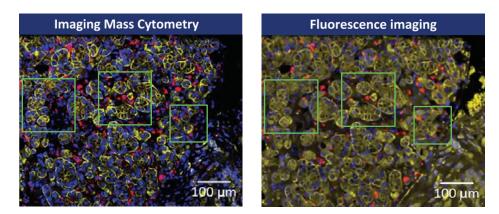
Whether you are analyzing suspension or tissue samples, time-of-flight (TOF) technology combined with Maxpar<sup>®</sup> reagents enables a streamlined end-to-end workflow to complete high-parameter experiments faster than fluorescence-based detection.

#### **Tissue imaging**

Get started with our Maxpar IMC<sup>™</sup> Cell Segmentation Kits and ready-to-go high-plex panels.



#### **Clear spatial imaging**



The Imaging Mass Cytometry<sup>™</sup> image (far left) shows many welldefined red signals from CD68 that are indistinct or missing from the fluorescence image (left).

#### Without compromise.

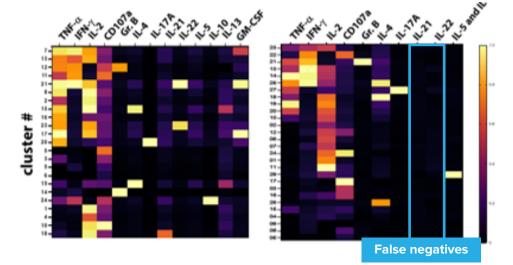
#### Results you can trust, reproduce and publish

## **See more** with CyTOF Flow cytometry

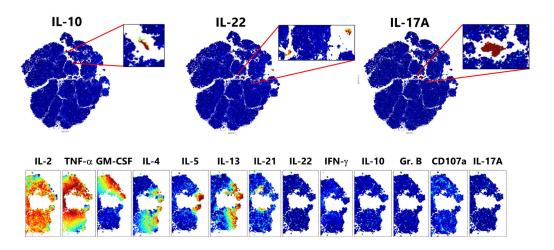
CyTOF detects more immune subpopulations in a single cell compared to full spectrum flow cytometry (FSFC).

#### Detect more cell populations with greater functional diversity.

The heat map on the left shows detection of IL-21 and IL-22, not present in the spectral data. Additionally, IL-5 and IL-10 were detected by CyTOF in independent channels.



#### Clear resolution and detection of low-frequency immune cells



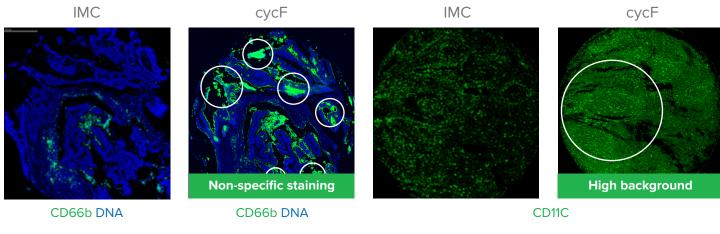
Application of opt-SNE to this CyTOF dataset highlights distinct combinations of effector functions from type 1, type 2 and type 17 lineages.

This data features findings from a comparison study using a 28-marker common panel including 12 cytokines. Data provided by Boston University.

### See your true biology

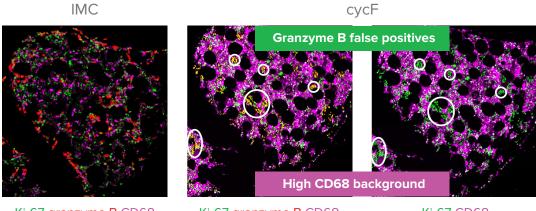
#### **See clearly** with IMC Tissue Imaging

IMC allows highly specific staining without the challenges of autofluorescence or false positives inherent to cyclic fluorescence-based imaging (cycF).



The cycF image (right) shows non-specific falsepositive CD66b signal in colon adenocarcinoma. CD11c signal is obscured by autofluorescence with cycF (right) in lymph node tissue.

#### Image highly-autofluorescent tissue types without challenge



In bone marrow, cycF data (right) shows granzyme B signal co-localized to the nucleus with Ki-67. The normal cytoplasmic or membranous localization of granzyme B can only be seen with IMC (left). CD68 signal is clear with IMC but obscured with cycF.

Ki-67 granzyme B CD68

Ki-67 granzyme B CD68

Ki-67 CD68

Data generated from a comparison study with multiplex cyclic fluorescence (cycF) using a 27-marker common panel. Cyclic fluorescence data provided by Georgetown University.

## The CyTOF Advantage.

A trusted technology used in

Over 200 clinical trials

Over 2,000 peer-reviewed publications

For **any** phase of research

#### Learn more: standardbio.com

#### For Research Use Only. Not for use in diagnostic procedures.

Information in this publication is subject to change without notice. Limited Use Label License: The purchase of this Standard BioTools Instrument and/or Consumable conveys to the purchaser the limited, nontransferable right to use only with Standard BioTools Consumables and/or Instruments respectively except as approved in writing by Standard BioTools Inc. (f.k.a. Fluidigm Corporation): www.standardbio.com/legal/salesterms. Patents: www.standardbio.com/legal/notices. Trademarks: Standard BioTools, the Standard BioTools logo, Fluidigm, the Fluidigm logo, CyTOF, Direct, Imaging Mass Cytometry, IMC, Immune Profiling Assay and Maxpar are trademarks and/or registered trademarks of Standard BioTools Inc. or its affiliates in the United States and/or other countries. All other trademarks are the sole property of their respective owners. ©2023 Standard BioTools Inc. All rights reserved. 05/2023

FLDM-01131 Rev 01