

Translating the complex immune system into disease insights with CyTOF technology



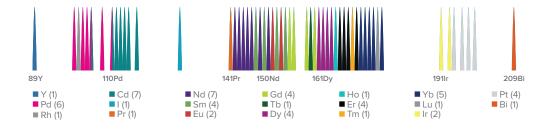
Reasons to choose CyTOF

THE CYTOF

Reasons to take your high-dimensional

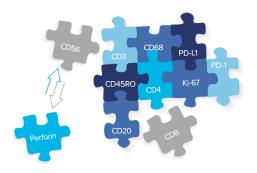
Precision data with discrete signals

Not impacted by spectral overlapping of fluorochromes and tissue autofluorescence



Easy panel design to complete experiments quicker

Large number of available antibodies without overlap simplifies panel design and expansion



Start with ready-to-go panels and easily swap markers in and out.

B

Reproducible and comparable

Stained samples can be frozen, stored and shipped to support longitudinal studies and multi-site workflows.

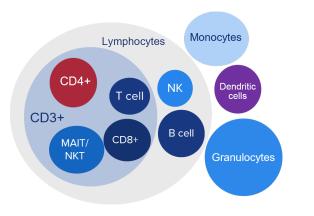


ADVANTAGE

research beyond the limits of fluorescence

Capture rare or unexpected cell populations

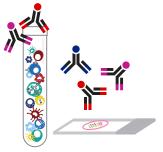
Unbiased, high-dimensional profiling of 40-plus markers to uncover diverse immune subpopulations





Minimal sample required, saving on limited clinical research material

Simultaneous staining and detection from a single tube or tissue scan, without multiple staining controls or time-consuming cyclic protocols



6

Trusted by researchers

The leading technology for high-parameter immune research





Cytometry by time-of-flight (CyTOF[®] technology)

Applies purified heavy metal labels, not normally found in biological systems, instead of fluorophores



Risks of fluorescence

for high-parameter studies



Missed cell populations or **false positives**



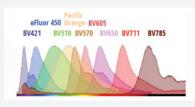




Reduced sensitivity where fluorescence overlap occurs



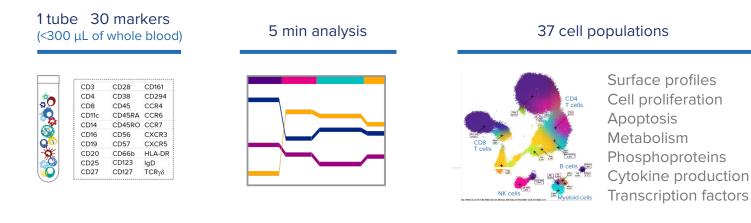
Higher resource use to compensate for spectral overlap



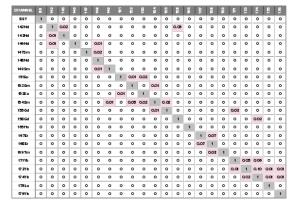
From sample collection to high-dimensional insights **in 3 days***

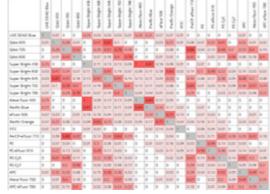
Flow cytometry

Get started with the validated Maxpar[®] Direct[™] Immune Profiling Assay[™].



Minimal signal overlap





The CyTOF flow cytometry image (far left) shows minimal spillover between metal channels when compared with the same panel from a competitor spectral flow cytometer (left).

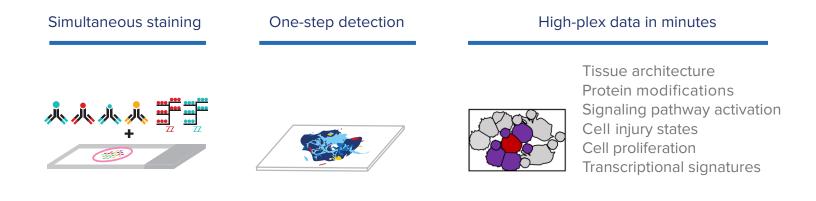
Quantify and visualize 40-plus markers in a single run.

* After panel and image analysis optimization

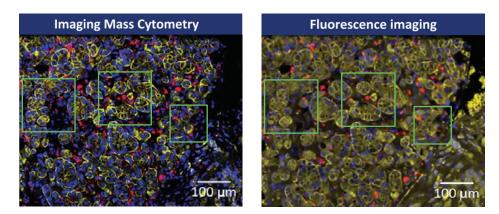
Whether you are analyzing suspension or tissue samples, time-of-flight (TOF) technology combined with Maxpar[®] reagents enables a streamlined end-to-end workflow to complete high-parameter experiments faster than fluorescence-based detection.

Tissue imaging

Get started with our Maxpar IMC[™] Cell Segmentation Kits and ready-to-go high-plex panels.



Clear spatial imaging



The Imaging Mass Cytometry[™] image (far left) shows many welldefined red signals from CD68 that are indistinct or missing from the fluorescence image (left).

Without compromise.

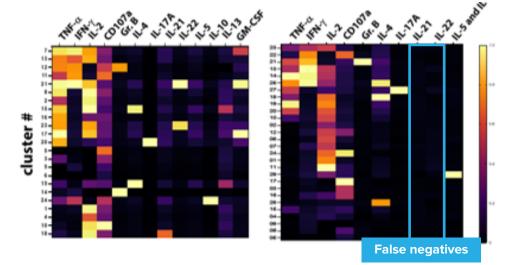
Results you can trust, reproduce and publish

See more with CyTOF Flow cytometry

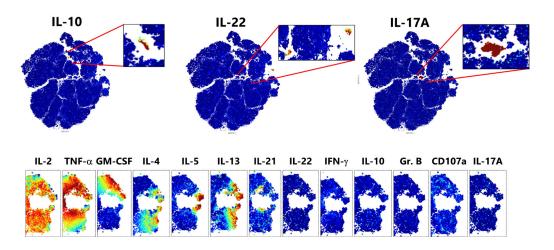
CyTOF detects more immune subpopulations in a single cell compared to full spectrum flow cytometry (FSFC).

Detect more cell populations with greater functional diversity.

The heat map on the left shows detection of IL-21 and IL-22, not present in the spectral data. Additionally, IL-5 and IL-10 were detected by CyTOF in independent channels.



Clear resolution and detection of low-frequency immune cells



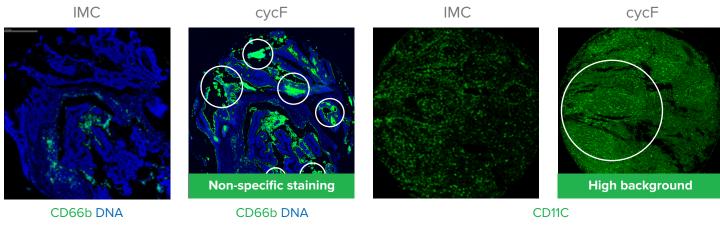
Application of opt-SNE to this CyTOF dataset highlights distinct combinations of effector functions from type 1, type 2 and type 17 lineages.

This data features findings from a comparison study using a 28-marker common panel including 12 cytokines. Data provided by Boston University.

See your true biology

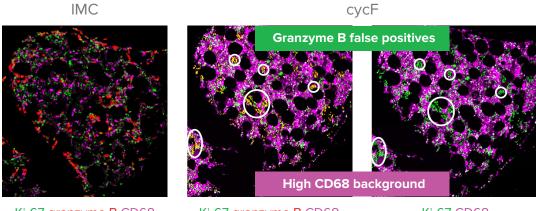
See clearly with IMC Tissue Imaging

IMC allows highly specific staining without the challenges of autofluorescence or false positives inherent to cyclic fluorescence-based imaging (cycF).



The cycF image (right) shows non-specific falsepositive CD66b signal in colon adenocarcinoma. CD11c signal is obscured by autofluorescence with cycF (right) in lymph node tissue.

Image highly-autofluorescent tissue types without challenge



In bone marrow, cycF data (right) shows granzyme B signal co-localized to the nucleus with Ki-67. The normal cytoplasmic or membranous localization of granzyme B can only be seen with IMC (left). CD68 signal is clear with IMC but obscured with cycF.

Ki-67 granzyme B CD68

Ki-67 granzyme B CD68

Ki-67 CD68

Data generated from a comparison study with multiplex cyclic fluorescence (cycF) using a 27-marker common panel. Cyclic fluorescence data provided by Georgetown University.

The CyTOF Advantage.

A trusted technology used in

Over 200 clinical trials

Over 2,000 peer-reviewed publications

For **any** phase of research

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